

# JC-1 MitoMP Detection Kit

## Technical Manual

Technical Manual (Japanese version) is available at <http://www.dojindo.co.jp/manual/MT09.pdf>

### General Information

Mitochondria is one of the important organelles in discussing early cytotoxicity, oxidative stress and apoptosis in vivo<sup>1)</sup>. Mitochondria synthesize ATP using oxygen to produce necessary energy for living cells. Lowering of mitochondrial activity and dysfunction are known to be closely related to cancer, aging, and neurodegenerative diseases such as Alzheimer's and Parkinson's diseases<sup>2), 3)</sup>.

JC-1 is widely used for observing mitochondrial membrane potential, shows fluorescence characteristic change from green (530 nm) to red (590 nm) depending on the mitochondrial membrane potential.

A red/green fluorescence intensity ratio of JC-1 decreases in depolarized mitochondria due to disruption of red fluorescent J-aggregates.

Unlike commercially available JC-1, no precipitation of JC-1 in aqueous buffer occurs by using this kit. In addition, the Imaging Buffer including in this kit minimizes fluorescence background and maintains the cells being healthy during assays.

### Kit Contents

JC-1 Dye 100 nmol x 1  
Imaging Buffer (10x) 11 ml x 1

### Storage Condition

Store at 0–5 °C.

### Required Equipment and Materials

- Dimethyl sulfoxide (DMSO)
- Medium or HBSS
- Micropipettes
- Microtube

Table 1 Required amount of Imaging Buffer solution by vessel type

Vessel	35 mmdish	ibidi 8 well plate
Appropriate amount	2 ml/dish	200 µl/well
Vessel	96 well plate	Sample tube (Flow Cytometry)
Appropriate amount	100 µl/well	0.5 ml/sample

### Preparation of Solutions

#### Preparation of Imaging Buffer solution

Dilute Imaging Buffer (10x) 10 times using double-deionized water.  
Use the Imaging Buffer solution within the day.

#### Preparation of JC-1 DMSO stock solution (2 mmol/l)

Add 50 µl of DMSO to a tube of JC-1 Dye and dissolve the red-purple solid by pipetting.

Store the JC-1 DMSO stock solution at -20 °C.

\*Protect the JC-1 DMSO stock solution from light.

\*JC-1 DMSO stock solution is stable at -20°C for up to one month.

#### Preparation of JC-1 working solution

Take a necessary amount of the JC-1 DMSO stock solution with a micropipette and transfer it to a microtube.

Add appropriate amount of medium to the tube, then mix immediately by 10 times of pipetting.

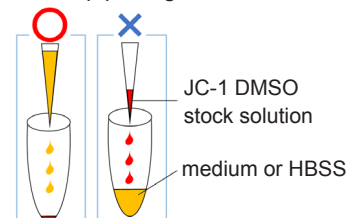
Adjust the final concentration of JC-1 in the range of 1–15 µmol/l.

\*To prepare a homogeneous working solution, please exactly follow the preparing procedure above.

\*Equilibrate both DMSO stock solution and culture medium to room temperature before preparing a working solution.

\*When the fluorescence signal is weak, optimize the concentration of JC-1 in working solution and incubation time.

\*Use the JC-1 working solution within the day.



### General protocol

#### Mitochondrial membrane potential determination using JC-1

1. Inoculate cells into a dish or a chamber slide and incubate the cells at 37 °C in a 5% CO<sub>2</sub> incubator.

2. Add an appropriate volume of JC-1 working solution to wells.\*<sup>1</sup>

3. Incubate the cells at 37 °C in the 5% CO<sub>2</sub> incubator for 30–60 minutes.

4. Discard the supernatant and wash the cells with medium or HBSS twice.\*<sup>2</sup>

5. Add Imaging Buffer solution and observe the cells under a fluorescence microscope.\*<sup>3</sup>

\*<sup>1</sup> Do not wash the cells after depolarization with CCCP or FCCP because the washing step might change mitochondrial membrane potential.

\*<sup>2</sup> Do not use PBS for washing to avoid a damage to the cells.

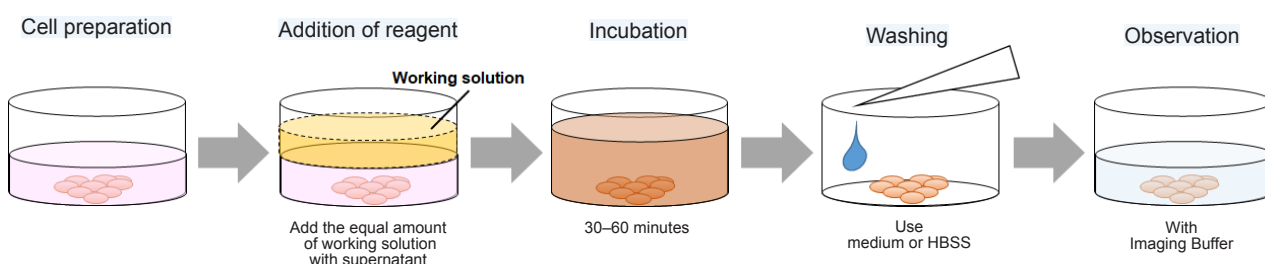
\*<sup>3</sup> Do not irradiate the excitation light to the sample for long time. JC-1 dye is light sensitive.

#### Example: Staining procedure with JC-1 (as 2 µmol/l of final concentration)

Transfer 2 µl of JC-1 DMSO stock solution (2 mmol/l) to a microtube.

Add 1 ml of medium to the tube, then mix immediately by 10 times of pipetting.

Add the equal amount of JC-1 working solution with supernatant to a well or dish.



### Fluorescence microscopic detection of mitochondrial membrane potential in HeLa cells treated with Carbonyl cyanide m-chlorophenyl hydrazone (CCCP)

1. HeLa cells ( $2.4 \times 10^5$  cells/ml, 200  $\mu$ l) in MEM (10% fetal bovine serum, 1% penicillin-streptomycin) were seeded on a  $\mu$ -slide 8 well plate (ibidi) and were cultured at 37 °C in a 5% CO<sub>2</sub> incubator overnight.
2. The medium was removed and CCCP in MEM (0 or 100  $\mu$ mol/l, 200  $\mu$ l) was added to each well, and then the cells were cultured at 37 °C for 90 minutes in the 5% CO<sub>2</sub> incubator.
3. After the supernatant (100  $\mu$ l) was removed, JC-1 working solution (4  $\mu$ mol/l, 100  $\mu$ l) was added, then the cells were cultured at 37 °C for 30 minutes in the 5% CO<sub>2</sub> incubator.
4. The cells were washed with 200  $\mu$ l of HBSS twice.
5. Imaging Buffer solution (200  $\mu$ l) was added, and the cells were observed under a fluorescence microscope.

Fluorescence imaging:

Green Ex: 488 nm, Em: 500–550 nm

Red Ex: 561 nm, Em: 560–610 nm

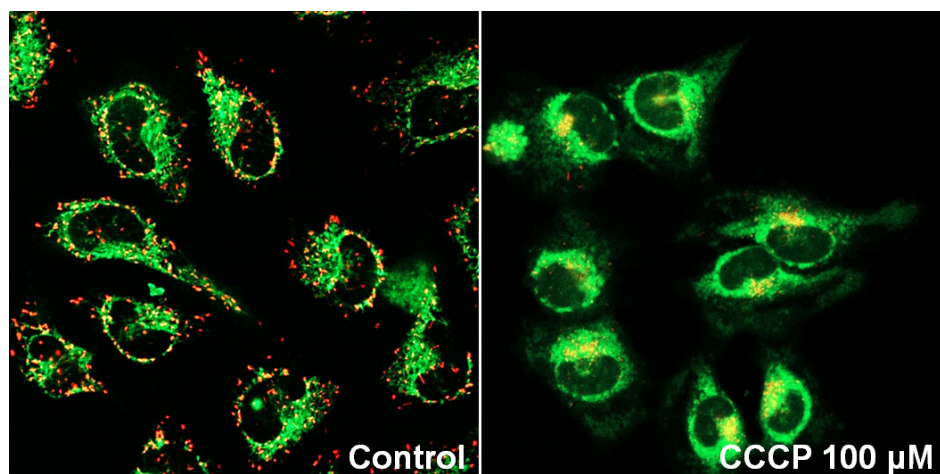


Figure 1 Fluorescence imaging of mitochondrial membrane potential in HeLa cells

### Fluorescence microscopic detection of mitochondrial membrane potential in HeLa cells treated with Carbonyl cyanide-p-trifluoromethoxyphenyl hydrazone (FCCP)

1. HeLa cells ( $2.4 \times 10^5$  cells/ml, 200  $\mu$ l) in MEM (10% fetal bovine serum, 1% penicillin-streptomycin) were seeded on a  $\mu$ -slide 8 well plate (ibidi) and were cultured at 37 °C in a 5% CO<sub>2</sub> incubator overnight.
2. The medium was removed and FCCP in MEM (0 or 100  $\mu$ mol/l, 200  $\mu$ l) was added to each well, and then the cells were cultured at 37 °C for 30 minutes in the 5% CO<sub>2</sub> incubator.
3. After the supernatant (100  $\mu$ l) was removed, JC-1 working solution (4  $\mu$ mol/l, 100  $\mu$ l) was added, then the cells were cultured at 37 °C for 30 minutes in the 5% CO<sub>2</sub> incubator.
4. The cells were washed with 200  $\mu$ l of HBSS twice.
5. Imaging Buffer solution (200  $\mu$ l) was added, and the cells were observed under a fluorescence microscope.

Fluorescence imaging:

Green Ex: 488 nm, Em: 500–550 nm

Red Ex: 561 nm, Em: 560–610 nm

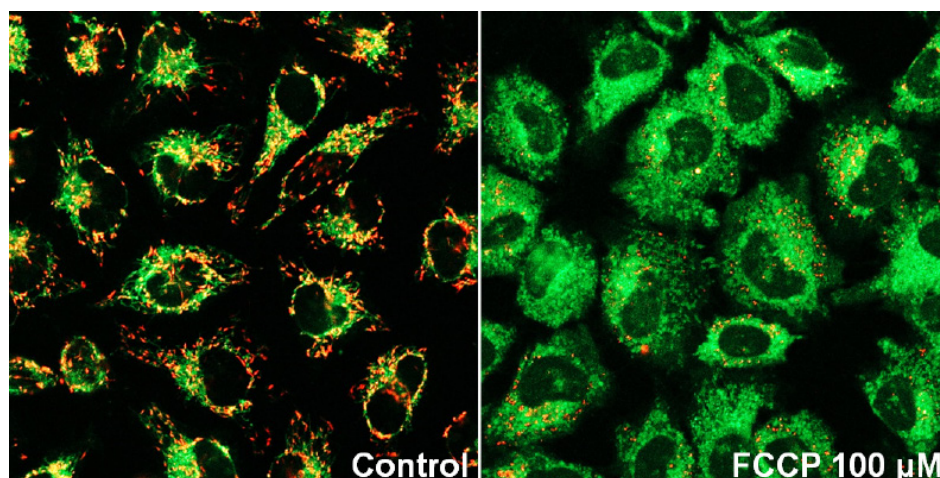


Figure 2 Fluorescence imaging of mitochondrial membrane potential in HeLa cells

### Mitochondrial membrane potential detection of apoptosis-induced Jurkat cells

1. Jurkat cells ( $1.0 \times 10^6$  cells/ml, 2 ml) in RPMI (10% fetal bovine serum, 1% penicillin-streptomycin) were transferred to a 5 mL tube.
2. The medium was removed and staurosporine in RPMI (0 or 2.5  $\mu\text{g/l}$ , 2 ml) was added to each tube, and then the cells were incubated at 37 °C for 150 minutes in a 5% CO<sub>2</sub> incubator.
3. JC-1 working solution (4  $\mu\text{mol/l}$ , 2 ml) was added, then the cells were incubated at 37 °C for 30 minutes in the 5% CO<sub>2</sub> incubator.
4. The media was removed and the cells were washed with 200  $\mu\text{l}$  of HBSS twice.
5. Imaging Buffer solution (2 ml) was added, and the cells were observed under a fluorescence microscope, and analyzed using a plate reader or a flow cytometer.

#### Fluorescence imaging

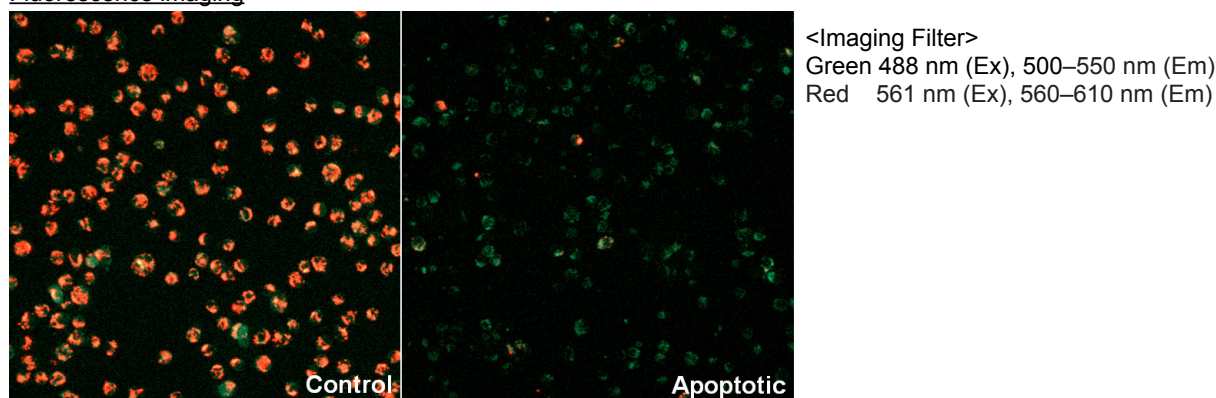


Figure 3 Fluorescence imaging of mitochondrial membrane potential in Jurkat cells

#### Plate Reader Detection

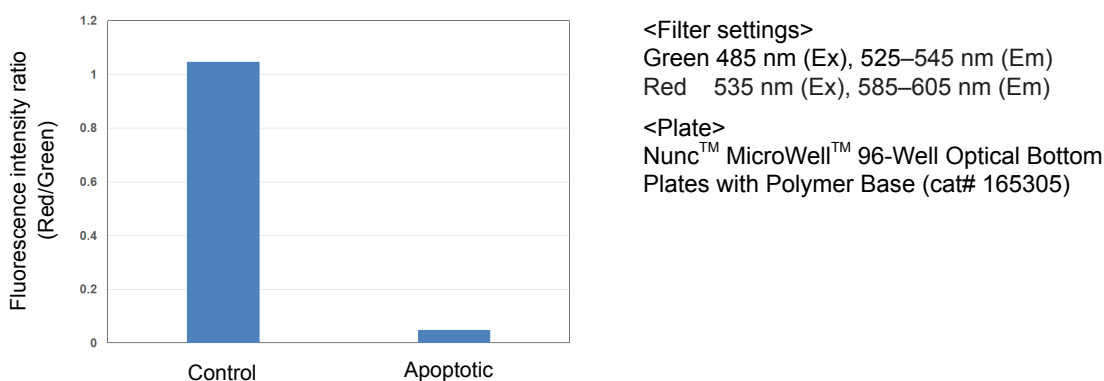


Figure 4 Fluorescence intensity ratio of mitochondrial membrane potential in Jurkat cells

#### Flow Cytometry

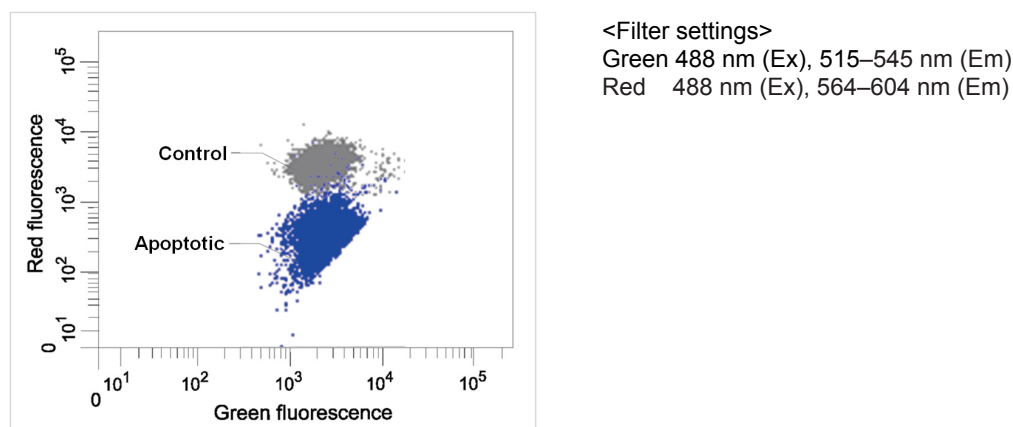


Figure 5 Flow cytometric analysis of mitochondrial membrane potential in Jurkat cells

### References

- 1) Ferri, K. F. et al., *J. Exp. Med.*, **2000**, 192, 1081–1092.
- 2) Matsuda, N. et al., *J. Cell Biol.*, **2010**, 189, 211.
- 3) Wang, J. L. et al., *PNAS*, **2000**, 97, 7124–7129.
- 4) Birju, P. S. et al., *ACS Nano*, **2014**, 8, 9, 9379–9387.
- 5) Cristiana, F. et al., *JID*, **2000**, 115, 5, 835–841.

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