Reagent for Cellular Function Analysis



Reagent for Cellular Function Analysis

Autophagy

- Autophagic Flux Assay Kit
- DALGreen-Autophagy Detection
- DAPGreen-Autophagy Detection
- DAPRed-Autopagy Detection

Senescence

- Cellular Senescence Detection Kit -SPiDER-&Gal
- Cellular Senescence Plate Assay Kit -SPiDER-βGal
- Cellular Senescence Detection Kit
 -SPiDER Blue

Neurodegenerative Diseases

Cancer

Senescence

Mitochondria

- Mitophagy Detection Kit
- JC-1 MitoMP Detection Kit
- MitoBright LT Series
- MT-1 MitoMP Detection Kit
- MitoBright ROS Deep Red
- Extracellular OCR Plate Assay Kit

Cellular Metabolism

- Glycolysis/OXPHOS Assay Kit
- ATP Assay Kit-Luminescence
- Lactate Assay Kit-WST

Ferroptosis **Example 2**

- FerroOrange
- Liperfluo
- Mito-FerroGreen
- MitoPeDPP
- Cystine Uptake Assay Kit
- MDA Assay Kit
- Lipid Peroxidation Probe
 -BDP 581/591 C11-

Cancer				
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MT09	JC-1 MitoMP Detection Kit	33		
MT10, MT11, MT12	MitoBright LT Series	37		
MT13	MT-1 MitoMP Detection Kit	33		
MT16	MtoBright ROS Deep Red - Mitochondrial Superoxide Detection	35		
MD01	Mitophagy Detection Kit	34		
MT02	Mtphagy Dye	34		
E297	Extracellular OCR Plate Assay Kit	32		
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D676	DAPGreen - Autophagy Detection	12		
D677	DAPRed - Autophagy Detection	12		
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L248	Liperfluo	16		
M489	Mito-FerroGreen	36		
UP05	Cystine Uptake Assay Kit	29		
G270	Glycolysis/OXPHOS Assay Kit	24		
A550	ATP Assay Kit-Luminescence	26		
L256	Lactate Assay Kit-WST	25		
R252	ROS Assay Kit -Highly Sensitive DCFH-DA-	14		
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	Neurodegenerative Diseases	
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SG05	Cellular Senescence Plate Assay Kit - SPiDER-βGal	11
MT09	JC-1 MitoMP Detection Kit	33
MT10, MT11, MT12	MitoBright LT Series	37
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MT16	MtoBright ROS Deep Red - Mitochondrial Superoxide Detection	35
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D675	DALGreen - Autophagy Detection	12
D676	DAPGreen - Autophagy Detection	12
D677	DAPRed - Autophagy Detection	12
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G270	Glycolysis/OXPHOS Assay Kit	24
A550	ATP Assay Kit-Luminescence	26
L256	Lactate Assay Kit-WST	25
R252	ROS Assay Kit -Highly Sensitive DCFH-DA-	14
R253	ROS Assay Kit -Photo-oxidation Resistant DCFH-DA-	15
G257	GSSG/GSH Quantification Kit	19
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LD05	Lipid Droplet Assay Kit - Blue	43
LD06	Lipid Droplet Assay Kit - Deep Red	43

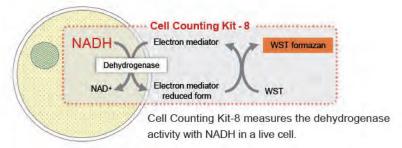
Cell Counting Kit-

Cell Proliferation / Cell Death

Cell Counting Kit-8

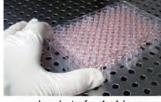


Detection Principle



Easy Step



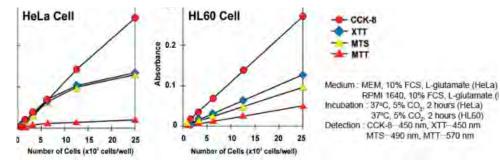




Add CCK-8 solution

Incubate for 1- 4 hours Measure O.D. at 450 nm

Sensitivity comparison



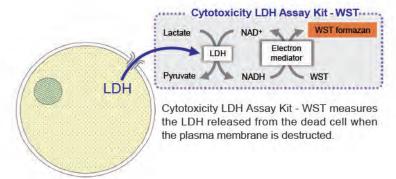
Description	Unit	Code
	500 tests	CK04-05
-8	1000 tests	CK04-11
8	3000 tests	CK04-13
	10000 tests	CK04-20

Cell Proliferation / Cell Death

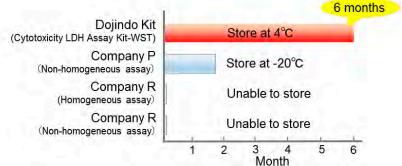
Cytotoxicity LDH Assay Kit-WST



Detection Principle



Stable Working Solution



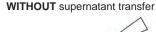
Working Solution is stable for 6 months under refrigerated conditions. Therefore, after the preparation, working solution can be used as a ready- to use solution at any time during this period.

Nearly 90% lower total cost* compared with short-stability LDH kits.

*Based on a comparison of 2000 tests over a continuous sixmonth usage period. Actual results may vary depending on assay frequency and local pricing.

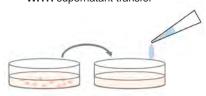
Choose between Two Procedures

Cytotoxicity LDH Assay Kit-WST can be applied with and without supernatant transferring. Please choose suitable method for your experiment.





WITH supernatant transfer



Descri	ption	Unit	Code
		100 tests	CK12-01
Cytotoxicity LDH Assay Kit-WST		500 tests	CK12-05
-,		2000 tests	CK12-20

Cell Proliferation / Cell Death

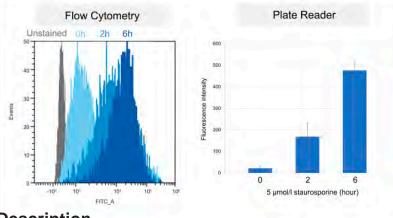
Annexin V Apoptosis Plate Assay Kit



Accurate plate assay without the need for washing

Typically, apoptotic cells are detected using flow cytometry or fluorescence microscopy, but these are time-consuming to process multiple samples. This kit contains fluorescently labelled Annexin V and a reagent (Quenching Buffer) that quenches the fluorescence of Annexin V not bound to PS, allowing rapid detection of multiple samples using a plate reader without the need for washing procedures.

Comparison with flow cytometry



[Experimental conditions]

Cell type: HeLa cells

Staurosporine concentration: 5 µmol/l

Time: 0 to 6 hours

[Detection conditions]

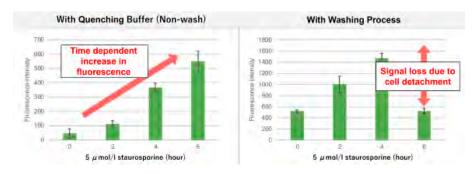
Plate reader: TECAN Infinite M200 PRO.

bottom reading

Flow cytometer: SONY SA3800

Description

Conventional plate-reader assays with washing steps can detach dying cells, leading to variability and reduced fluorescence. This kit combines FITC-labelled Annexin V, which binds PS, with a quenching buffer, eliminating washing. The resulting no-wash PS assay enables simple, reliable and accurate detection of apoptosis.



Description Unit Code

Cell Proliferation / Cell Death

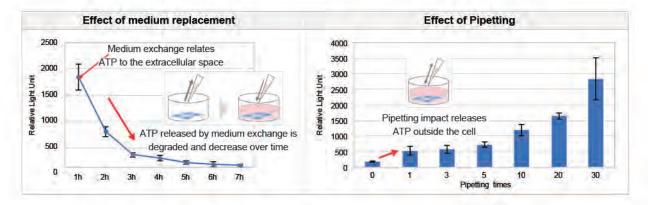
Extracellular ATP Assay Kit-Luminescence



Accurate measurement protocol

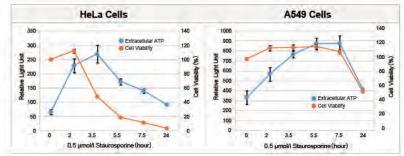
Extracellular ATP, one of the indicators of dead cells, can be measured.

ATP release into the extracellular space is highly sensitive and can occur even after medium changes or pipetting. This kit allows first-time users to measure extracellular ATP changes due to stimulation using the luciferase luminescence method, with a protocol that accounts for measurement influences.



Experimental Example: Evaluation using Staurosporine-treated cells

HeLa and A549 cells were treated with Staurosporine, and extracellular ATP release and cell viability were assessed over time. The results showed that the extracellular ATP release of each cell differed in amount and timing depending on the cell type.



[Product in use]

Extracellular ATP : Extracellular ATP Assay Kit-

Luminescence (code: E299)

Cell viability: Cell Counting Kit-8 (code: CK04)

Description	Unit	Code
Extracellular ATP Assay Kit-Luminescence	100 tests	E299-10

SG03 SG07 Senescence Detection

Cellular Senescence Detection Kit - SPiDER-\(\beta \) Gal Cellular Senescence Detection Kit - SPiDER Blue

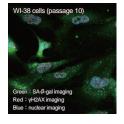


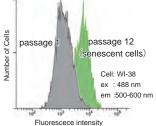


Cellular Senescence Detection Kit – SPiDER-βGal allows to detect SA-β-gal with high sensitivity and ease of use. SPiDER-βGal is a new reagent to detect β-galactosidase which possesses a high cell-permeability and a high retentivity inside cells. SA-β-gal are detected specifically not only in living cells but also fixed cells by using a reagent (Bafilomycin A1) to inhibit endogenous β-galactosidase activity. Therefore, SPiDER-βGal can be applied to quantitative analysis by flow cytometry.

SPiDER-Boal

Compatible with quantitative analysis





Compatible Instruments: ✓ Microscope ✓ Flow Cytometer ✓ Plate Reader

X-Gal

Difficult to quantify

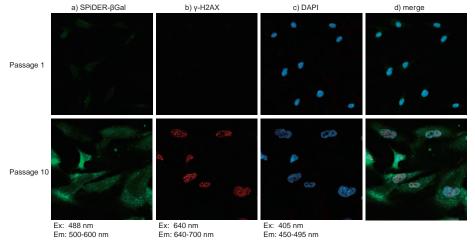


Difficult to... · count the cells · difficult to distinguish /

negative cells

Compatible Instruments: ✓ Microscope

Experimental Example: Co-staining of SA- β-gal and DNA Damage marker



WI-38 cells were treated with anti- v-H2AX antibody and observed under a confocal microscope. The procedure involved several steps, including fixing the cells, permeabilizing, blocking, adding primary and secondary antibodies, staining with DAPI, and washing the cells. The experiment aimed to detect DNA damage and study DNA repair pathways.

Description	Unit*	Code
Cellular Senescence Detection Kit - SPiDER-βGal	10 assays	SG03-10
Cellular Senescence Detection Kit - SPiDER Blue	1 Plate	SG07-10

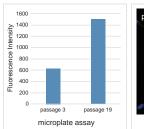
Proliferation Cell death

Cellular Senescence Plate Assay Kit - SPiDER-βGal



This kit allows you to quantify SA-β-gal activity and evaluate multiple samples in a 96-well plate by simply adding SPiDER-βGal, a reagent that can detect β-galactosidase.

Correlation with Imaging Data



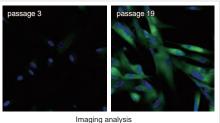


Plate Assav Ex. 535nm / Em. 580nm

Imaging data

Green: Ex. 488nm / Em. 500-600nm (SA-β-Gal staining with Cellular Senescence Detection Kit – SPiDER-βGal(Code SG04)) Blue: Ex. 405nm / Em. 450-495nm (Nuclear staining with -Cellstain- DAPI solution(Code D523))

Experimental Procedure



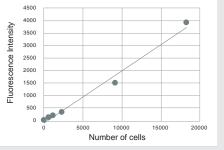
Cell Count Normalization Kit

Combined Cellular Senescence Plate Assay protocol available online



When normalized to the results obtained by quantifying nucleic acids using the Cell Count Normalization Kit, the measured values of SA-βgal activity become available for evaluation of SA-β-gal activity according to cell number.

Highly correlated to cell number



Description	Unit	Code
Cellular Senescence Plate Assay Kit - SPiDER-βGal	20 tests	SG05-01
Cellulal Sellescelice Flate Assay Kit - SFIDER-pGal	100 tests	SG05-05
Cell Count Normalization Kit	200 tests	C544-02
Cell Court Normalization Kit	1000 tests	C544-10

Endocytosis

etc

Autophagy D675

DAPGreen / Red - Autophagy Detection DALGreen - Autophagy Detection



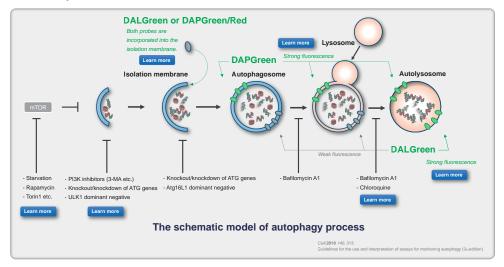


D676



D677

DAPGreen and DAPRed detect autophagosomes, while DALGreen detects autolysosomes. These dyes are permeable to cells and enables live cell imaging with fluorescence microscopy, and DAPGreen and DALGreen allow for quantitative assay by flow cytometry. Autophagy is an intracellular degradation system involving autophagosome formation, detected by DAPGreen and DAPRed, and lysosome fusion, detected by DALGreen, which fluoresces intensity increases in acidic conditions.



Feature of Each Dye

	Fluorescent	licable instrum	Microplate	Fluorescent properties	Volume / the number of usable assays	Existing methods
DAPGreen	Microscope	cytometer	reader	Ex = 425-475 nm Em = 500-560 nm * For confocal microscope, the sample can be excited at 488 nm	5 nmol x 1 / 35 mm dish: 25 (when used in 1.0 µmol/l)	LC3-GFP
DAPRed	0	×	×	Ex = 500-560 nm Em = 690-750 nm	5 nmol x 1 / 35 mm dish: 25 (when used in 1.0 µmol/l)	MDC Cyto-ID etc.
DALGreen	0	0	×	Ex = 350-450 nm Em = 500-560 nm * For confocal microscope, the sample can be excited at 488 nm	20 nmol x 1 / 35 mm dish: 10 (when used in 1.0 µmol/l)	LC3-GFP-RFP etc.

^{*}Double staining imaging by DAPGreen and DALGreen is not possible

Autophagic Flux Kit



Senescence

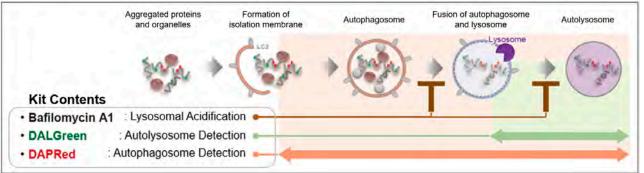
Proliferation Cell death

Autophagy

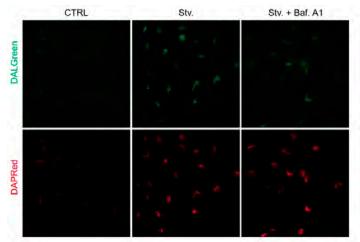
Oxidative Stress

Metabolism

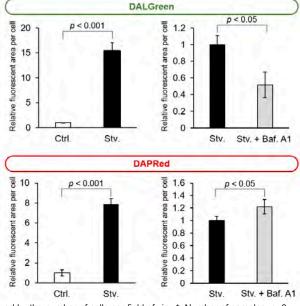
Aggregated proteins and organelles



Experimental Example: Autophagy Flux Analysis



By culturing HeLa cells in HBSS with starvation, autophagy was induced and DAPRed and DALGreen fluorescence increased. Addition of Baf. A1 decreased DALGreen fluorescence, indicating that autolysosomes were reduced and Autophagy Flux was inhibited.



Quantification method: Fluorescence values (area) were obtained in Image J and normalized by the number of cells per field of view*. Number of samples: n=3 *Please obtain images with the same number of cells per field of view as possible.

Description	Unit*	Code
Autophagic Flux Assay Kit	1 set	A562-10
DALGreen - Autophagy Detection	20 nmol	D675-10
DAPGreen - Autophagy Detection	5 nmol	D676-10
DAPRed - Autophagy Detection	5 nmol	D677-10

Other Organelles

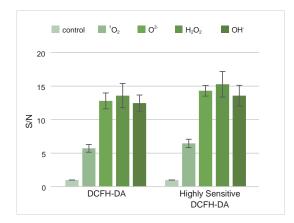
Oxidative Stress

ROS Assay Kit -Highly Sensitive DCFH-DA-



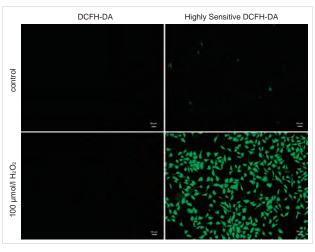
The selectivity for ROS

DCFH-DA is widely used for ROS detection, but it has some limitations like weak fluorescence signals and high background. Dojindo's ROS Assay Kit -Highly Sensitive DCFH-DA- allows ROS detection with higher sensitivity than DCFH-DA with the similar ROS selectivity.

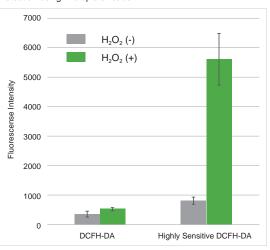


Experimental Example: High Sensitive Detection Compared with DCFH-DA









Hydrogen peroxide (H₂O₂)-treated HeLa cells (1 × 10⁴ cells/ml) were stained with DCFH-DA or the ROS Assay Kit-Highly Sensitive DCFH-DA, and the fluorescence intensity of intracellular ROS was compared between two detection kits. As a result, the ROS Assay Kit-Highly Sensitive DCFH-DA in high-sensitivity detection of intracellular ROS was better than DCFH-DA.

Description	Unit	Code
ROS Assay Kit -Highly Sensitive DCFH-DA-	100 tests	R252-10

Proliferation Cell death

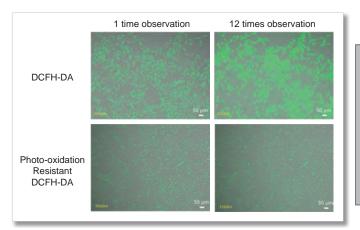
Senescence

Autophagy

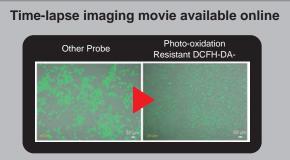
ROS Assay Kit -Photo-oxidation Resistant DCFH-DA-

The dye used in this kit allows ROS detection with higher sensitivity than DCFH-DA and long-term observation of live cells due to its resistance to photooxidation.

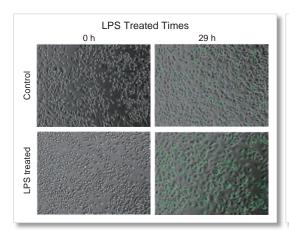
Resistant to Photo-oxidation

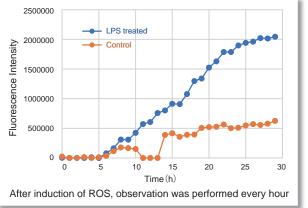


Comparison of photo-oxidation resistant ability in HeLa cells



Experimental Example: Simultaneous Detection of ROS in LPS-treated macrophages





In Lipopolysaccharide (LPS) treated RAW 264.7 cells, after being stained with regular DCFH-DA, Highly Sensitive DCFH-DA, or Photo-oxidation Resistant DCFH-DA, the intracellular ROS level was compared.

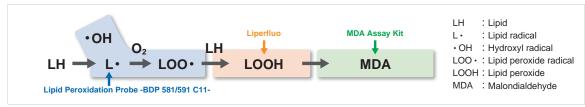
Description	Unit	Code
ROS Assay Kit -Photo-oxidation Resistant DCFH-DA-	100 tests	R253-10

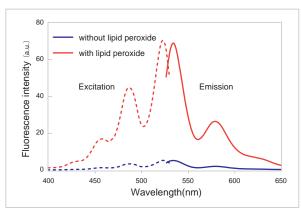
Lipid Peroxide Detection

Liperfluo



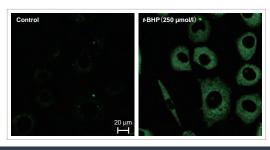
Liperfluo is a Dojindo-developed fluorescence probe to specifically detect lipid peroxides with minimal photo-damage or auto-fluorescence. It emits intense fluorescence in organic solvents and is nearly non-fluorescent in aqueous media. Liperfluo's tetraethyleneglycol group increases its solubility and makes it suitable for imaging lipid peroxides in cell membranes. It's used to monitor lipid peroxidation in ferroptosis research through fluorescence microscopy and flow cytometry.





Excitation and emission without lipid peroxide spectra of Liperfluo with or without lipid peroxide in ethanol.

Experimental Example: Lipid Peroxide Detection in Living Cells



Liperfluo added to cells, t-BHP induced lipid peroxidation and cells were observed under confocal microscope to study ferroptosis.

Cell line: L929

Microscope: Zeiss LSM510META

Filter type: FITC (GFP, Alexa488) wide filter

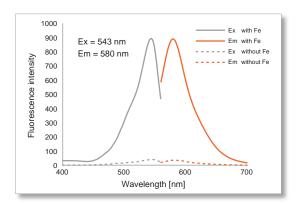
HFT UV/488 NFT490 BP505-550

	Description	Unit*	Code
Liperfluo		1 set (50 μ g $ imes$ 5)	L248-10

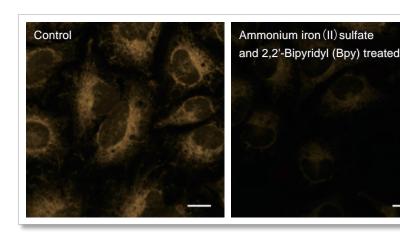
Intracellular Iron Ion Measurement

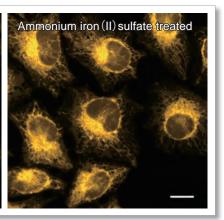
FerroOrange

FerroOrange is a specific probe for ferrous ions (Fe²⁺) and reacts irreversibly, which differs from the detection principle of chelating ability. Fluorescence imaging or quantification of intracellular Fe²⁺ is often used to evaluate iron-related non-apoptotic cell death, ferroptosis.



Experimental Example





HeLa cells treated with chelator of iron 2,2'-bipyridyl (Bpy) (100 μ mol/l) or Ammonium iron (II) sulfate (100 μ mol/l) were prepared. The change of intracellular Fe²⁺ in HeLa cells was detected by the FerroOrange. Ex = 561 nm, Em = 570-620 nm, Scale bars 20 μ m

	Description	Unit*	Code
FerroOrange		1 tube 3 tube	F374-10 F374-12

Live-cell imaging of copper (I) ions

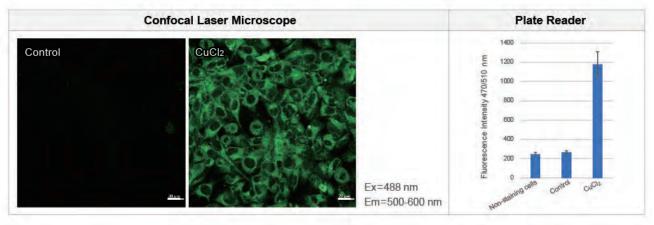
CuprosGreen



CuprosGreen is a fluorescent probe that selectively reacts with intracellular copper (I) ions and emits fluorescence. Its membrane permeability allows live-cell imaging of copper (I) ions.

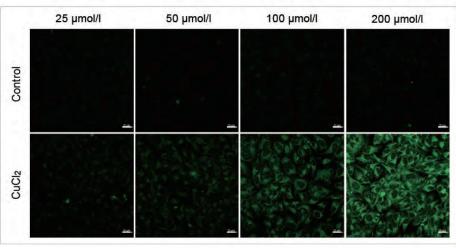
Plate reader assays are also possible.

Microscopic observation and plate reader detection possible



Experimental Example

We evaluated concentration-dependent changes in intracellular CuCl₂ levels in HeLa cells and observed a concentration-dependent increase in intracellular CuCl₂ levels upon CuCl₂ addition compared to control by confocal microscopy.

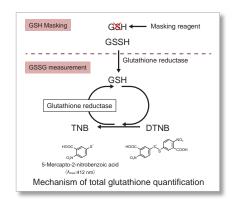


	Description	Unit	Code
CuprosGreen		20 μΙ	C557-10

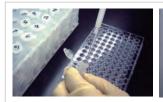
Quantification of Reduced (GSH) and Oxidized (GSSG) Glutathione GSSG/GSH Quantification Kit



The GSSG/GSH Quantification kit contains Masking Reagent of GSH. GSH will be deactivated in the sample by simply adding the Masking Reagent. Then, using the enzymatic recycling system, only the GSSG will be detected by measuring the absorbance (λ max = 412 nm) of DTNB (5,5-dithio-bis- (2-nitrobenzoic acid). The quantity of GSH can also be determined, by substracting GSSG from the total amount of glutathione. With this kit, GSH/GSSG concentrations from 0.5 μ mol/l to 50 μ mol/l and GSSG concentrations from 0.5 μ mol/l can be quantified.



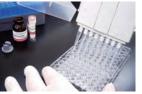
Experimental Procedure: Experimental time is only 1-2 hours



- GSSG/GSH Standard Solution and add Sample A or Sample B to each well.
- 2) Add Buffer solution to each well



Incubate at 37°C for 1 h.

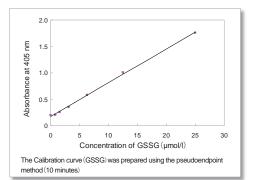


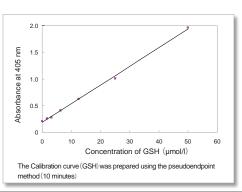
4)-5) Add substrate working solution and Enzyme/ Coenzyme working solution to each well.



6)-7) After incubating at 37°C for 10 minutes, measure the absorbane of each well with a microplate

Calibration Curve





Description	Unit	Code
GSSG/GSH Quantification Kit	200 tests	G257-10

Lysosome

Related Products

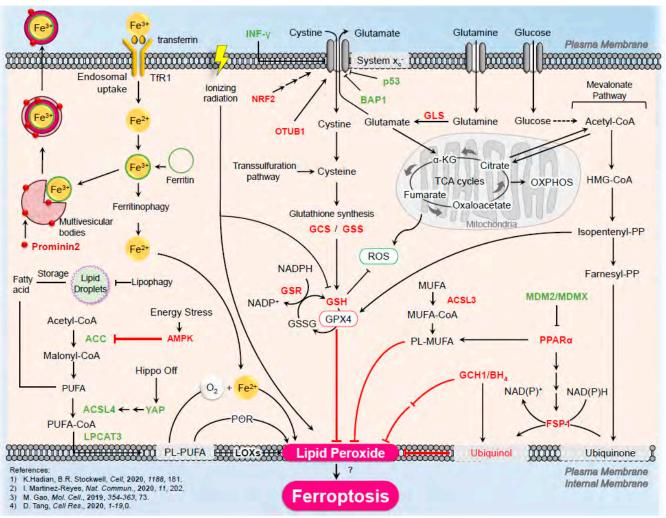
Ferroptosis research



"Ferroptosis" was coined by Stockwell et al. at Columbia University in 2012 and described as a form of irondependent cell death. * It was reported to be a form of programmed cell death by the Nomenclature Committee on Cell Death (NCCD) in 2018.

Ferroptosis is a form of programmed cell death caused by iron ion-dependent accumulation of lipid peroxides. Ferroptosis has been shown to follow a different cell death pathway from apoptosis and thus is attracting attention as a new target for cancer therapy. It has also been found to be associated with various diseases, such as neurodegenerative diseases, cerebral apoplexy, and hepatitis (NASH).

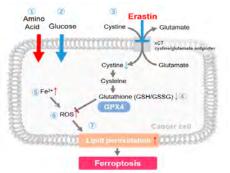
Ferroptosis Pathway

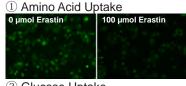


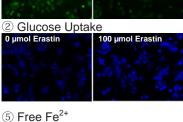
Experimental Example: Evaluating Intracellular Uptake and Redox Balance

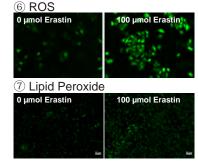
0 µmol Erastin

We investigated the transition of cellular metabolisms in A549 cells treated with Erastin, a known ferroptosis inducer. Our results revealed the following.









- Cystine Uptake 4 Redox Balance 700 500 0.50 400 300 0.40 200 100 0 µmol 100 µmol 0 µmol 100 µmol . Erastin Erastin Erastin Erastin
- Experimenta Condition

100 µmol Erastin

- <Experimenta Condition> A549 cells were treated with 100 μmol/l Erastin/MEM for 3 hours.
- The inhibition of cystine uptake by Erastin led to a depletion of cysteine, which in turn increased the compensatory uptake of other amino acids.
- Glucose uptake, which typically promotes ferroptosis, was found to decrease upon Erastin treatment, suggesting a potential cellular self-defense mechanism.
- The depletion of cysteine resulted in a decrease in glutathione levels and an increase in Fe²⁺, ROS, and lipid peroxides, all of which are recognized markers of ferroptosis.

Description	Unit	Code
Liperfluo (Used in Experiment ⑦)	50 μg x 5	L248-10
Lipid Peroxidation Probe -BDP 581/591 C11-	200 tests	L267-10
FerroOrange (Used in Experiment ⑤)	1 tube 3 tubes	F374-10 F374-12
Mito-FerroGreen	1 set (50 µg x 2)	M489-10
ROS Assay Kit -Highly Sensitive DCFH-DA- (Used in Experiment ⑥)	100 tests	R252-10
GSSG/GSH Quantification Kit (Used in Experiment ④)	200 tests	G257-10
MDA Assay Kit	100 tests	M496-10
Amino Acid Uptake Assay Kit (Used in Experiment ①)	20 tests 100 tests	UP04-10 UP04-12
Glucose Uptake Assay Kit (Used in Experiment ②)	1 set	UP01-10 (Blue) UP02-10 (Green) UP03-10 (Red)
Cystine Uptake Assay Kit (Used in Experiment ③)	20 tests 100 tests	UP05-10 UP05-12
Glycolysis/OXPHOS Assay Kit	50 tests	G270-10
Lyso-FerroRed	35 nmol	L270-10
Iron Assay Kit - Colorimetric -	50 tests	I291-10

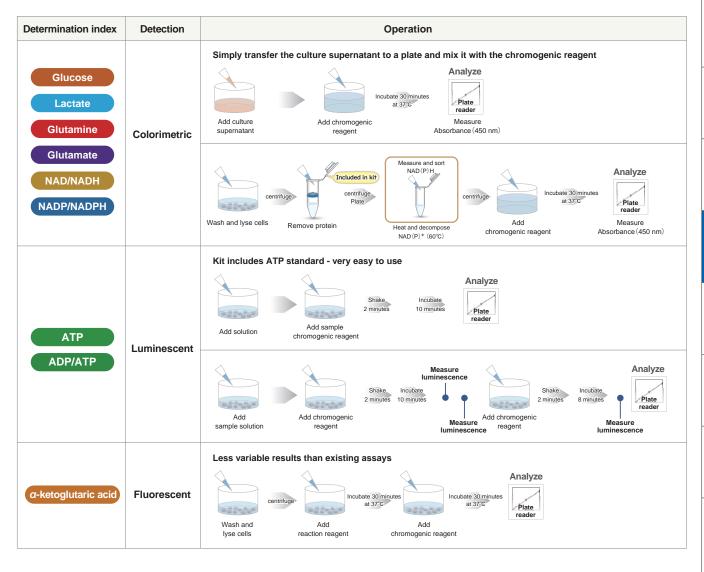
Measurements of Intracellular Metabolism



Description	Unit	Code
arter Kit		
Glycolysis/OXPHOS Assay Kit	50 tests	G270-10
Glycolysis/JC-1 MitoMP Assay Kit	50 tests	G272-10
uantification for Intracellular Metabolism		
ATD Account to Lumpin accounts	50 tests	A550-10
ATP Assay Kit-Luminescence	200 tests	A550-12
ADP/ATP Ratio Assay Kit-Luminescence	100 tests	A552-10
Change Assert Wit MCT	50 tests	G264-05
Glucose Assay Kit-WST	200 tests	G264-20
Glutamine Assay Kit-WST	100 tests	G268-10
Glutamate Assay Kit-WST	100 tests	G269-10
α-Ketoglutarate Assay Kit-Fluorometric	100 tests	K261-10
Lastata Assa KUWOT	50 tests	L256-10
Lactate Assay Kit-WST	200 tests	L256-20
NAD/NADH Assay Kit-WST	100 tests	N509-10
NADP/NADPH Assay Kit-WST	100 tests	N510-10
otake Assay Kit		
Glucose Uptake Assay Kit-Blue	1 set	UP01-10
Glucose Uptake Assay Kit-Green	1 set	UP02-10
Glucose Uptake Assay Kit-Red	1 set	UP03-10
Assistant Asial Hatalian Assass	20 tests	UP04-10
Amino Acid Uptake Assay	100 tests	UP04-12
Overline Harteles Assess Kit	20 tests	UP05-10
Cystine Uptake Assay Kit	100 tests	UP05-12
Fatty Acid Uptake Assay Kit	100 tests	UP07-10

Simple Procedure for First Time User

For a first-time user, the kit includes the reagents and components necessary for measuring samples. You'll soon realize how easy it is to use.



Intracellular Metabolism

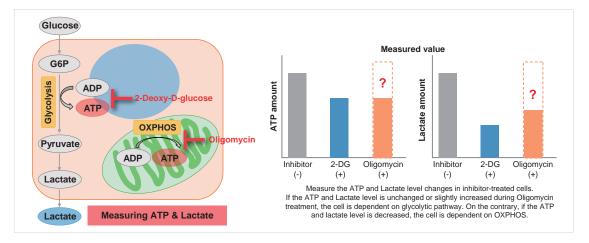
Glycolysis/OXPHOS Assay Kit



- Easy test via plate reader, no need for expensive equipment
- All reagent acquired is included, ready to use kit
- Easy-to-understand detailed protocol

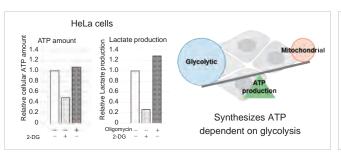
Combining methods (1) and (2) can be used to measure the metabolic pathway dependency of cells.

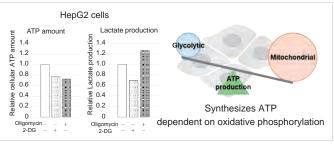
Cells are treated with oligomycin or 2-DG to inhibit OXPHOS or ATP synthesis in the glycolytic pathway, and the amounts of ATP and lactate production are measured, respectively. Changes in the amount of ATP can be used to determine the efficiency of energy production, and changes in the amount of lactate produced can be used to determine changes in glycolytic capacity and evaluate whether cells are dependent on glycolysis or OXPHOS.



Experimental Example:

Comparison of metabolic pathway dependence in different cell line





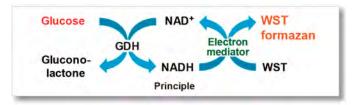
Description	Unit	Code
Glycolysis/OXPHOS Assay Kit	50 tests	G270-10

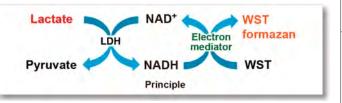
G264

Mitochondria

Glucose Assay Kit-WST Lactate Assay Kit-WST

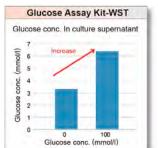
The Glucose Assay Kit-WST and Lactate Assay Kit-WST are colourimetric kits for the quantification of glucose and lactate, respectively, both with a lower detection limit of 0.02 mmol/L. These two indicators are crucial for understanding glycolytic metabolism and are among the most commonly measured parameters in metabolic studies. In addition, the use of Dojindo's proprietary WST dye in the assay systems allows for easy and highly accurate detection.

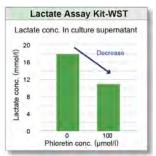




Experimental Example: Evaluation of culture supernatant using two indicators

Phloretin, the glucose transporter inhibitor, was added to Jurkat cells and the intracellular metabolism change was evaluated using Glucose Assay Kit-WST and Lactate Assay Kit-WST.

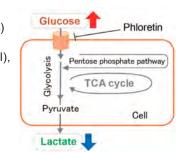




<Experimental condition>
Cell Line: Jurkat cells (5x10⁵ cells)
Stimulation condition:
Phloretin (final conc.: 100 μmol/l),

overnight incuvation

Sample: Culture supernatant"



<Description>

Glucose consumtion has decreased due to inhibition of glucose uptake by Phloretin, resulting in increase of glucose and decrease in lactate in culture supernatant.

Description	Unit	Code
Glucose Assay Kit-WST	50 tests 200 tests	G264-05 G264-20
Lactate Assay Kit-WST	50 tests 200 tests	L256-10 L256-20

Oxidative Stress

Intracellular Metabolism

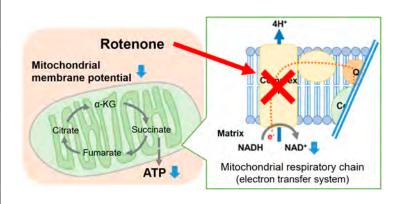
ATP Assay Kit-Luminescence

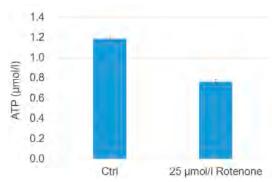


ATP Assay Kit-Luminescence is a luciferase luminescence assay kit for quantification of intracellular ATP. ATP is an important energy source of living cells that is synthesized in both glycolysis and mitochondrial oxidative phosphorylation. Mitochondrial dysfunction reduces ATP levels in cells and the decreased ATP levels are known to be associated with cancer, aging, and neurodegenerative diseases. Therefore, ATP level is used as an indicator for mitochondrial activity. ATP level is a lso focused in Cancer research since the recent studies have revealed that although cancer cells were known to rely on glycolysis for ATP synthesis, a shift from glycolysis to oxidative phosphorylation occurs when glycolysis is suppressed.

Experimental Example: Change in intracellular metabolism of rotenone-treated cells

Rotenone, which is known to inhibit the mitochondrial electron transport chain, was added to Jurkat cells, followed by measurement of intracellular ATP using the ATP Assay Kit-Luminescence. As a result, ATP production in the mitochondrial respiratory chain (the electron transport chain) was inhibited, and ATP concentrations were lower than those in the control cells.



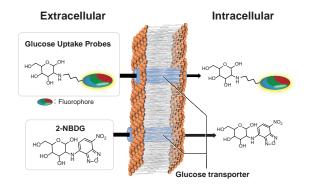


]	escription	Unit	Code
ATP Assay Kit-Luminescence		50 tests	A550-10
		200 tests	A550-12

Glucose Uptake Assay Kit

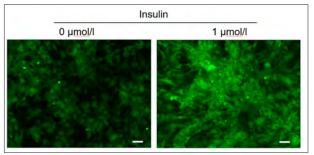
- Highly sensitive and simple measurement of glucose uptake capacity
- Applicable for microscopy & FCM
- Reduces dye leakage after staining

Principle



Experimental Example

Glucose uptake enhancement by insulin



Detailed results and other data are available online.

Comparison with Existing Method

The comparison of the Glucose Uptake Probe Series and the existing method(2-NBDG) is as below.

product name	Fluorescence microscope	Plate reader detection	FCM detection	Retention ability	Fluorescence characteristics
Glucose Uptake Assay Kit-Blue	0	×	0	1 hour *	λεx:386 nm λεm:474 nm
Glucose Uptake Assay Kit-Green	0	0	0	1 hour *	λex:507 nm λem:518 nm
Glucose Uptake Assay Kit-Red	0	0	0	1 hour *	λex:560 nm λem:572 nm
2-NBDG	0	×	0	30 minutes or less *	λex:465 nm λem:540 nm

*Result of A549 cells, the retention time for other cell lines may be different.

Description	Unit*	Code
Glucose Uptake Assay Kit-Blue	1 set	UP01-10
Glucose Uptake Assay Kit-Green	1 set	UP02-10
Glucose Uptake Assay Kit-Red	1 set	UP03-10

Oxidative Stress

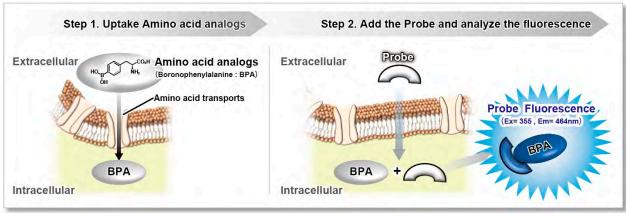
Intracellular Metabolism

Amino Acid Uptake Assay Kit



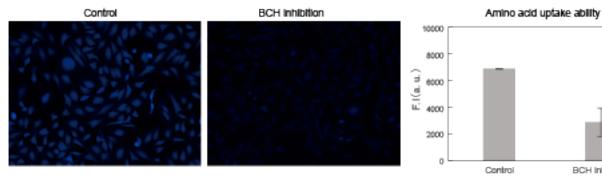
BCH inhibition

Principle



After amino acid analogs (BPA) are taken up into cells via amino acid transporters, the fluorescent probe permeates the cell membrane and binds to the amino acid analogs, emitting fluorescence

Experimental Example: Evaluation of BCH (Inhibitor of amino acids transporter)



Experiment Condiitons Cell Line: HeLa cells

Medium: MEM (5.5 mmol/l Glucose)

Incubation: 1 mmol/l BCH/HBSS (Hanks' Balanced Salt Solution), 37°C, 30 min Instrument: Fluorescent Microscopy (Ex=340-380 nm, Em: 435-485 nm)

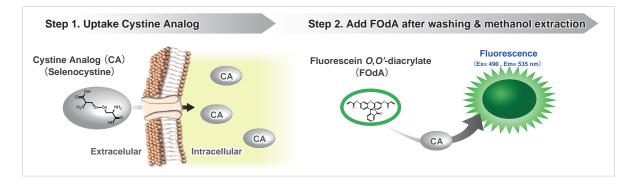
Instrument: Plate Reader (Ex=360 nm. Em: 460 nm)

Description	Unit	Code
·	20 tests	UP04-10
Amino Acid Uptake Assay Kit	100 tests	UP04-12

Cystine Uptake Assay Kit

- Easier way to cystine uptake assay
- Applied for plate assay

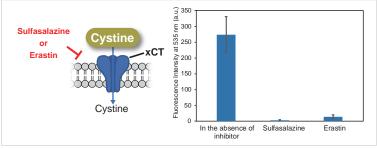
Principle



The Cystine Analog (CA) in this kit can be taken up into cells via xCT, and the incorporated CA can be specifically detected using the Fluorescent Probe and Reducing Agent. Thus, the xCT activity can be measured easily. [Patent applied]

Experimental Example: Evaluation of xCT Inhibitor Sulfasalazine or Erastin

Using this kit, we measured the inhibitory effect of sulfasalazine and erastin on cystine uptake by HeLa cells. The fluorescence intensity of the sulfasalazine and elastin groups decreased significantly, indicating that both reagents inhibit cystine uptake.



Experiment Condiitons

Cell Line: HeLa cells

Pretreatment: DMEM (cystine-free, serum-free), 37°C, 5 min Uptake conditions: 0.5 mmol/l sulfasalazine or 2 µmol/l erastin / Cystine Analog / DMEM (cystine-free, serum-free), 37°C, 30 min

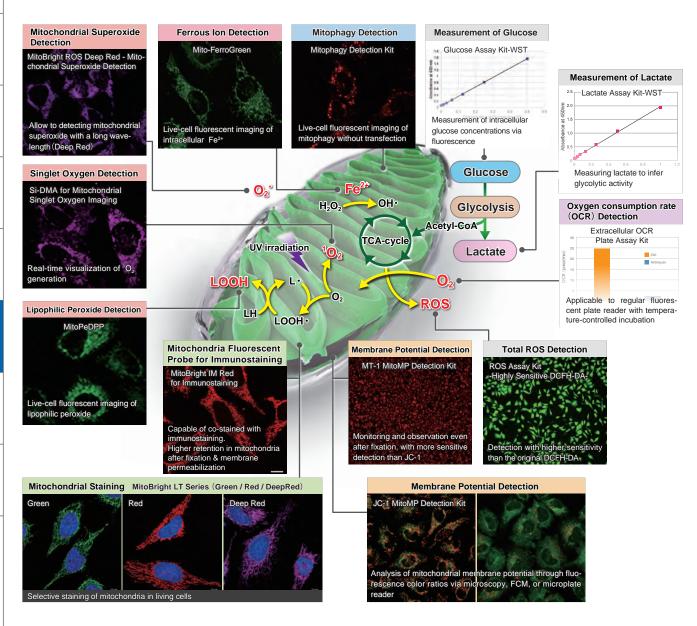
Instrument: Fluorescent Plate Reader Filter: Ex=485 nm, Em=535 nm

Description	Unit	Code
Cystine Uptake Assay Kit	20 tests 100 tests	UP05-10 UP05-12

Metabolism

Mitochondrial Research





Description	Unit	Code
Extracellular OCB Ploto Access Vit	100 tests	E297-10
Extracellular OCR Plate Assay Kit	300 tests	E297-12
Internal Index Occurs Detection Kit	100 tests	I306-10
Intracellular Oxygen Detection Kit	300 tests	1306-12
Chappe Association WCT	50 tests	G264-05
Glucose Assay Kit-WST	200 tests	G264-20
actate Assay Kit-WST	50 tests	L256-10
Lactate Assay Kit-WST	200 tests	L256-20
MT-1 MitoMP Detection Kit	1 set	MT13-10
JC-1 MitoMP Detection Kit	1 set	MT09-10
Mitophagy Detection Kit	1 set	MD01-10
Mtphagy Dye	5 μg × 3	MT02-10
MitoBright LT Green	400 μΙ	MT10-12
MitoBright LT Red	400 μΙ	MT11-12
MitoBright LT Deep Red	400 μΙ	MT12-12
Mita Dright IM Dad for Inspectorate in in a	20 µl×1	MT15-10
MitoBright IM Red for Immunostaining	20 μl×3	MT15-12
Mita Daialet DOO Danis Dadi. Mita akan 1821 O anni 112 Dati. 12	100 nmol × 1	MT16-10
MitoBright ROS Deep Red - Mitochondrial Superoxide Detection	100 nmol × 3	MT16-12
Mito-FerroGreen	1 set (50 μg × 2)	M489-10
Si-DMA for Mitochondrial Singlet Oxygen Imaging	2 µg	MT05-10
MitoPeDPP	5 μg × 3	M466-10

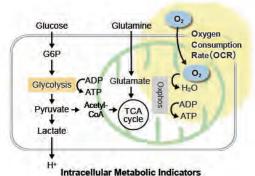
Mitochondrial Research

Extracellular OCR Plate Assay Kit Intracellular Oxygen Detection Kit





OCR is an Important Indicator for Mitochondrial Function



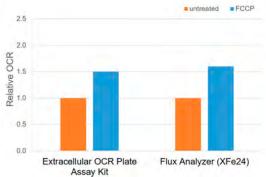
ATP production is oxygen-dependent; therefore, OCR is a key indicator in mitochondrial research.

Extracellular OCR Plate Assay Kit and Intracellular Oxygen Detection Kit provide an easy-to-use solution for your study.

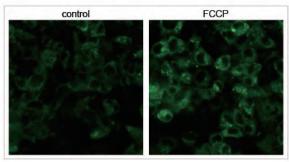
Comparison with Flux Analyzer (E297)

Correlation data of oxygen consumption rate changes were obtained for XFe24 and this kit.

Cells: HepG2
Cell Number: 5×10⁴ cells/well
Stimulation: FCCP
FCCP Concentration: 2 umol/l



Imaging analysis with fluorescence microscope (I306)



[Detection conditions] Cells: HepG2 Stimulation: FCCP

Confocal fluorescence microscope (ZEISS, LSM800)

Ex/Em = 488/550 - 700 nm

Laser: 1.0%, 700V Lens: ×40 Scan speed: 3

Description	Unit	Code
Extracellular OCR Plate Assay Kit	100 tests 300 tests	E297-10 E297-12
Intracellular Oxygen Detection Kit	100 tests 300 tests	1306-10 1306-12

MT09

Proliferation Cell death

Senescence

Autophagy

Oxidative Stress

Metabolism Mitochondria

JC-1 MitoMP Detection Kit MT-1 MitoMP Detection Kit

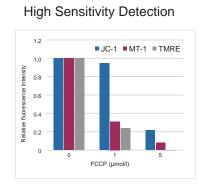
JC-1 indicates mitochondrial condition by changing from red to green fluorescence as mitochondrial membrane potential (MMP) decreases. While JC-1 and similar dyes such as TMRE and TMRM are popular for MMP detection, they suffer from low photostability and poor retention. Dojindo's MT-1 MitoMP Detection Kit overcomes these limitations and improves experimental reproducibility.

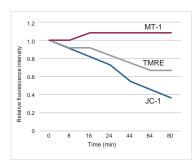
Comparison of Reagents

	Features	Sensitivity	Fixation	Monitoring	Fluorescence change (upon loss of mitochondrial membrane potential)	Detection (ex/em)
JC-1 (JC-1 MitoMP Detection Kit)	Recomended for starting-up	√			Color change from red to green	Green: 450-490 nm / 500-550 nm Red: 530-560 nm / 570-640 nm
MT-1 (MT-1 MitoMP Detection Kit)	Recommended for more detailed analysis	✓ (High)	√	1	Decrease in fluorescence intensity	530-560 nm / 570-640 nm
TMRE	Widely used	✓ (High)			Decrease in fluorescence intensity	530-560 nm / 570-640 nm

Experimental Example

JC-1 Detection of MMP in Apoptotic Cells





Allow to monitor MMP

Description	Unit*	Code
JC-1 MitoMP Detection Kit	1 set	MT09-10
MT-1 MitoMP Detection Kit	1 set	MT13-10

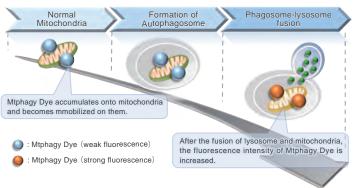
MT-1

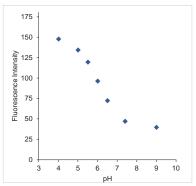
Mitochondrial Research

Mitophagy Detection Kit



This kit is composed of Mtphagy Dye, reagent for detection of mitophagy, and Lyso Dye. Mtphagy Dye accumulates in intact mitochondria, is immobilized on it with chemical bond and exhibits a weak fluorescence from the influence of surrounding condition. When Mitophagy is induced, the damaged mitochondria fuses to lysosome and then Mtphagy Dye emits a high fluorescence. To confirm the fusion of Mtphagy Dye—labeled mitochondria and lysosome, Lyso Dye included in this kit can be used.





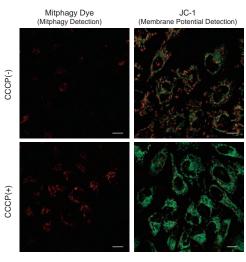
The fluorescent intensity of Mtphagy Dye is incresased at pH 4-5.

Experimental Example

Mitophagy Induction and Mitochondrial Membrane Potential Changes

Mitochondrial condition in the carbonyl cyanide m-chlorophenyl hydrazine (CCCP) treated Parkin-expressing HeLa cells was compared with untreated cells using Mitophagy Detection Kit (MD01, MT02) and JC-1 MitoMP Detection Kit (MT09).

As a result, mitophagy was hardly detected in the CCCP-untreated cells, and mitochondrial membrane potential was maintained normally. On the other hand, in CCCP-treated cells, we observed a decrease in mitochondrial membrane potential (decrease in red fluorescence of JC-1) and induction of mitophagy (increase in fluorescence of Mtphagy Dye).



Description	Unit*	Code
Mitophagy Detection Kit	1 set	MD01-10
Mtphagy Dye	5 μg × 3	MT02-10

Mitochondrial Superoxide Detection

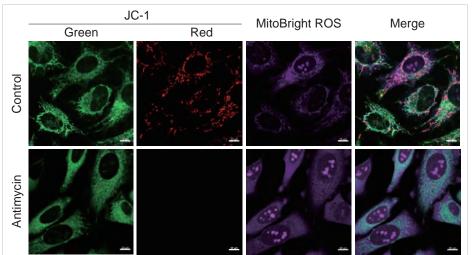
MitoBright ROS Deep Red - Mitochondrial Superoxide Detection



This dye emits deep red fluorescence; its fluorescence does not overlap with emission wavelengths that other red fluorescent markers use. Furthermore, the MitoBright ROS Deep Red is better able to selectively detect superoxide, compared to Company T's product Red.

Experimental Example

Simultaneously Evaluation of Mitochondrial Superoxide and Membrane Potential



<Imaging Conditions>
(Confocal microscopy)
JC-1: Green Ex = 488, Em = 490-520 nm,
Red: Ex = 561, Em = 560-600 nm
MitoBright ROS Deep Red: Ex = 633 nm,
Em = 640-700 nm
Scale bar: 10 μm

After HeLa cells were washed with HBSS, co-stained with MitoBright ROS Deep Red and mitochondrial membrane potential staining dye (JC-1: code MT09), and the generated mitochondrial ROS and membrane potential were observed simultaneously. As a result, the decrease in mitochondrial membrane potential and the generation of mitochondrial ROS are simultaneously observed.



Description	Unit*	Code
MitoBright ROS Deep Red - Mitochondrial Superoxide Detection	$100 \text{ nmol} \times 1$ $100 \text{ nmol} \times 3$	MT16-10 MT16-12

Mitochondrial Superoxide Detection

Mito-FerroGreen



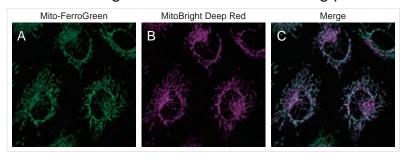


Mito-FerroGreen is a novel fluorescent probe for the detection of ferrous ion (Fe^{2+}) in mitochondria. Mito-FerroGreen has no chelating ability. Mito-FerroGreen and Fe^{2+} react irreversibly, which is different from the detection principle of calcium-iron probes such as Fluo-3.

* Bpy: 2,2' -Bipyridyl

Experimental Example

Double staining with mitochondrial staining probe



Double staining with mitocondrial staining probe

Mito-FerroGreen (5 µmol/l) Ex/Em = 488 nm/ 500-550 nm

MitoBright Deep Red (200 nmol/l) Ex/Em = 640 nm/ 656-700 nm

A Mito-FerroGreen

B MitoBright Deep Red

C Merge

HeLa cells incubated with Mito-FerroGreen and MitoBright Deep Red, treated with ammonium iron(II) sulfate, were observed by fluorescence microscopy.

Selection Guide of Iron Detection Dyes

	Mito-FerroGreen (M489)	FerroOrange (F374)
Localization	Mitochondria	Intracellular
Fluorescent Property	λex 505 nm, λem 535 nm	λex 543 nm, λem 580 nm
Instrument (filter)	Fluorescence microscope (FITC, GFP)	Fluorescence microscope, plate reader (Cy3)
Sample	Live Cell	Live cell
The number of assays	1 set (50 μg x 2) 10 assays at 35 mm dish (final concentration 5 μmol/l)	1 tube (24 μg) 17 assays at 35 mm dish (final concentration 1 μmol/l)

	Description	Unit*	Code
Mito-FerroGreen		1 set (50 μ g $ imes$ 2)	M489-10
FerroOrange		1 tube 3 tube	F374-10 F374-12

^{*}Approximate usage depends on the experiment. Please refer to our product webpage.

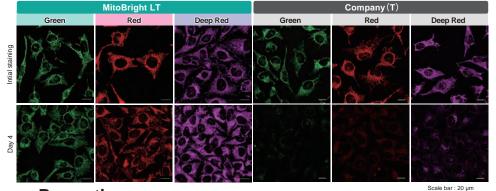
Mitochondrial Staining MitoBright LT Series



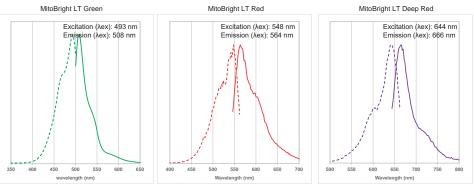
MitoBright LT dyes are designed to exhibit mitochondria retention for long-term visualization. In addition, the MitoBright LT dyes show stronger fluorescence signals compared with other commercially available dyes that contain the chloromethyl moiety. The MitoBright LT dyes offer three different color options (Green, Red and Deep Red), and are provided as a ready-to-use DMSO solution.

Experimental Example: Stained in serum-contained media

HeLa cells were stained with MitoBright LTs or an existing reagent and observed after 4 days. MitoBright LT remained unchanged and observable even after 4 days, while the existing reagent's intensity decreased.



Fluorescence Properties



Description	Unit*	Code
MitoBright LT Green	400 µl	MT10-12
MitoBright LT Red	400 µl	MT11-12
MitoBright LT Deep Red	400 µl	MT12-12

Oxidative Stress

Lysosomal Analysis

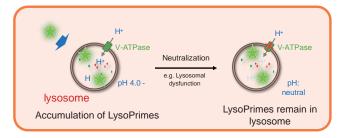
LysoPrime Green / Deep Red - High Specificity and pH Resistance



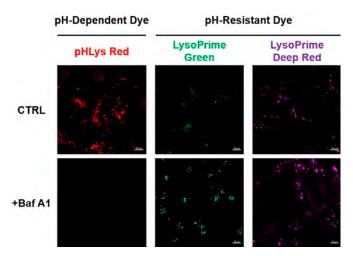


pHLys Red - Lysosomal Acidic pH Detection

pH-resistant Probe LysoPrime Green / Deep Red

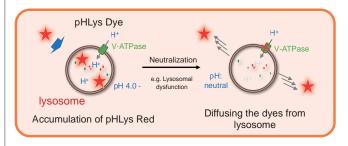


Resistance to pH changes

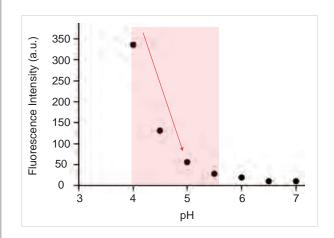


pHLys Red is highly specific to lysosomes and shows pH-dependent changes in fluorescence, and pH-resistant LysoPrime Green/Deep Red is retained in lysosomes even after adding Bafilomycin A1, a lysosomal acidity inhibitor. The lysosomal pH and mass of the same sample can be measured using these two dyes for a detailed analysis of lysosomal function.

pH-dependent Probe pHLys Red



pH dependence of pHLys Red



The fluorescence intensity of pHLys Red at each pH was confirmed in vitro, and it was confirmed that the fluorescence intensity changed sensitively within the range of lysosomal pH (pH 4.0-5.5).

Lysosomal Acidic pH Detection Kit





Senescence

Proliferation Cytotoxicity

Autophagy

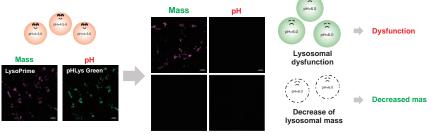
Oxidative Stress

Metabolism

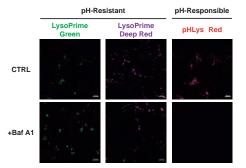
Mitochondria

The kit includes lysosome staining dyes, pHLys Red/Green (pH dependent), and LysoPrime Green/Deep Red (pH-independent). The pHLys and LysoPrime dyes accumulate in the intact lysosomes. The fluorescence intensity of pHLys dyes are enhanced as the acidity increases, and weak fluorescence is observed when lysosomes are neutralized due to the lysosomal dysfunction. On the other hand, LysoPrime dyes gives stable emissions even lysosomes are neutralized. Lysosomal pH and lysosomal mass can be measured by combining these pHLys and

LysoPrime dyes.

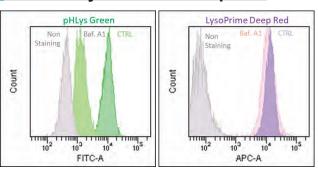


Imaging Analysis: Green/Red (#L266-10)



<Experimental Conditions> LysoPrime Green: Ex = 488 nm, Em = 490 – 550 nm pHLvs Red: Ex = 561 nm. Em = 560 – 620 nm

FCM Analysis: Green/Deep Red (#L268-10)



<Experimental Conditions>
pHLys Green: FITC Filter (Ex = 488 nm, Em = 515 – 545 nm)
LysoPrime Deep Red: APC Filter (Ex = 640 nm, Em = 650 – 670 nm)

Description	Unit*	Code
Lysosomal Acidic pH Detection Kit – Green/Red *1	1 set	L266-10
Lysosomal Acidic pH Detection Kit – Green/Deep Red *2	1 set	L268-10
LysoPrime Green – High Specificity and pH Resistance	10 μl × 1 10 μl × 3	L261-10 L261-12
LysoPrime Deep Red - High Specificity and pH Resistance	1 tube 3 tube	L264-10 L264-12
pHLys Red - Lysosomal Acidic pH Detection	1 tube 3 tube	L265-10 L265-12

^{*1} Green/Red: combination of LysoPrime Green and pHLys Red, *2 Green/Deep Red: combination of pHLys Green and LysoPrime Deep Red
*Approximate usage depends on the experiment. Please refer to our product webpage.

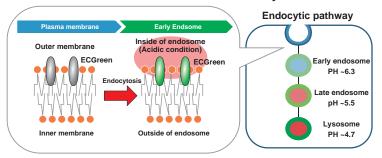
Endocytosis

ECGreen-Endocytosis Detection



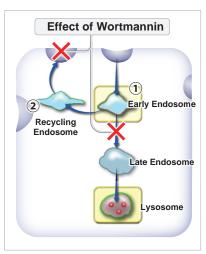
ECGreen-Endocytosis Detection is a pH dependent fluorescence dye that localizes to vesicle membrane. The visualization of endocytosis using the ECGreen is a more direct method than fluorescent analogs and allows visualization endocytosis from the stage of early endosomes.

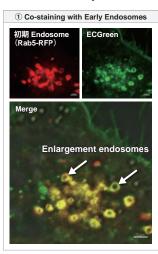
The detection mechanism of endocytosis

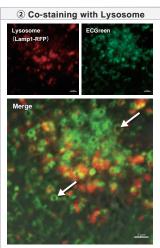


Clear visualization of intracellular vesicular trafficking

It has been known that Wortmannin inhibits the recycling of endosomes or transition to lysosomes and causes enlargement of endosomes. To evaluate these changes caused by Wortmannin, early endosomes were co-stained by ECGreen and Rab5-RFP (marker protein of early endosomes), and lysosomes were co-stained by ECGreen and lysosome staining reagent. In adding Wortmannin, ECGreen was colocalized with enlarged endosomes (Rab5-RFP). On the other hand, ECGreen wasn't colocalized with lysosomes.







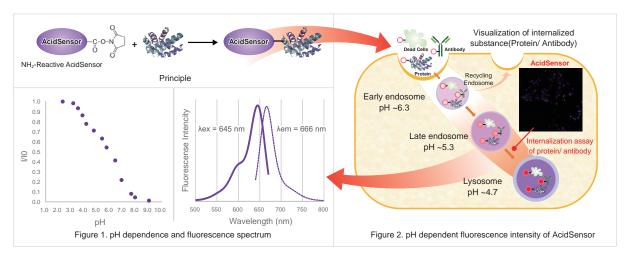
Description	Unit*	Code
ECGreen-Endocytosis Detection	40 µl	E296-10

AcidSensor Labeling Kit – Endocytic Internalization Assay

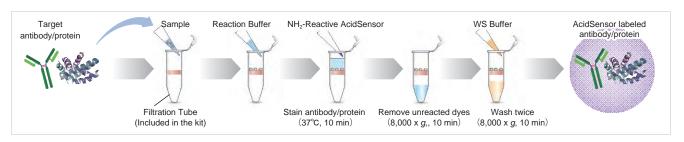


- a a

This kit is an all-in-one kit that allows visualization of the endocytosis uptake of a target substance. The NH₂-Reactive AcidSensor (fluorescent probe) included in the kit has an intramolecular active ester group that forms a stable covalent bond when mixed with an amino group-containing target substance (protein). The AcidSensor label can be excited at 633 nm, allowing for multiple staining with green or red fluorescence (Figure 1). The AcidSensor label shows little fluorescence in neutral conditions and fluoresces when acidified in the cells where it is taken up by endocytosis (Figure 2).



This kit includes a filtration tube necessary to remove the unreacted dye, and allows you to perform everything from labeling to purification operations.* In addition, even first-time users can easily label AcidSensor by conducting experiments according to the instruction manual. * Protein/Antibody is not included.



Description	Unit	Code
AcidSensor Labeling Kit – Endocytic Internalization Assay	3 samples	A558-10

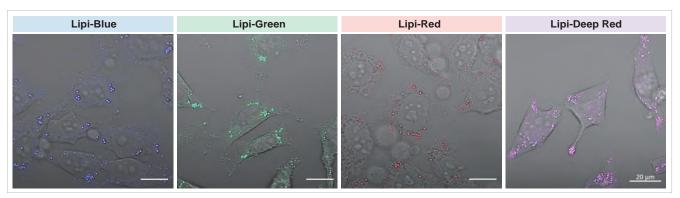
Oxidative Stress

Lipid Droplet Staining

Lipi-Blue / Green / Red / Deep Red



Lipi probes are small molecules that emit strong fluorescence in a hydrophobic environment such as LDs, which can be observed without any washing steps after staining with Lipi probes.



A medium that contained oleic acid (200 μ mol/l) was added and incubated overnight. Then, the supernatant was removed and the cells were washed with PBS. Each Lipi product series (1 μ mol/l) was added and the cells were incubated for 15 minutes.

Lipi-Blue: Ex. 405 nm / Em. 450 – 500 nm, Lipi-Green: Ex. 488 nm / Em. 500 – 550 nm, Lipi-Red: Ex. 561 nm / Em. 565 – 650 nm, Lipi-Deep Red: Ex. 640 nm / Em. 650-700 nm

Comparison of Reagents

	Dojindo		Othe		ther Products	
	Lipi-Blue	Lipi-Green	Lipi-Red	Oil Red O	Nile Red	Reagent B
Live Cells	✓	✓	✓		✓	✓
Fixed Cells	✓	✓	1	✓	✓	✓
Selectivity towards Lipid Droplet (Level of Background)	✓	✓	1			
General Filter Accommondation*1	✓	✓	✓* ²	n.d.	*3	✓
Retention in Live Cells	✓	1		n.d.		

^{*1} Please refer to our website for the co-staining filter.

 $^{^{13}}$ Leaks in GFP filter (500 \sim 540 nm)

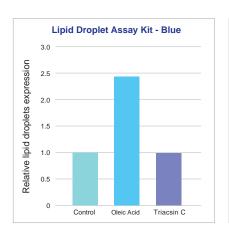
	Description Unit*	Code
Lipi-Blue	10 nmol $ imes$ 1	LD01-10
Lipi-Green	10 nmol × 1	LD02-10
Lipi-Red	100 nmol × 1	LD03-10
Lipi-Deep Red	10 nmol × 1	LD04-10

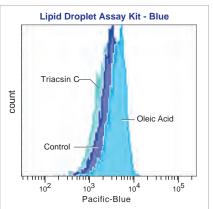
² When co-staining with a green fluorescent dye, a green fluorescent emission filter less than 550 nm is recommended.

Lipid Droplet Assay Kit - Blue / Deep Red

The Lipid Droplet Assay Kit simplifies the quantification of fat droplets with provided protocols and buffers. It works for both live and fixed cells. Compared to colorimetric reagents, it reduces measuring time and increases experiment repeatability by avoiding dye deposition in the plate.

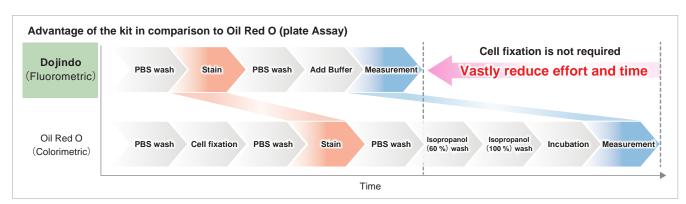
Experimental Example of plate assay and flow cytometry





Changes in lipid droplets were examined after the addition of oleic acid or Triacsin C (acyl-CoA synthetase inhibitor) to the A549 cell culture medium.

Advantage of the kit in comparison to Oil Red O (Plate Assay)



Description	Unit*	Code
Lipid Droplet Assay Kit-Blue	1 set	LD05-10
Lipid Droplet Assay Kit-Deep Red	1 set	LD06-10

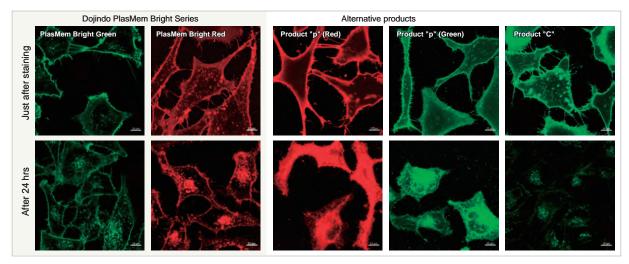
Cell Membrane Staining

PlasMem Bright Green / Red

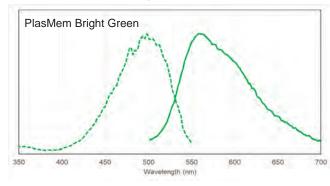


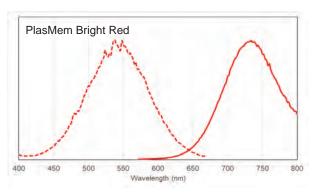
PlasMem Bright dyes are designed to stain plasma membrane for over a day. Furthermore, the PlasMem Bright dyes are more water-soluble compared with other commercially available dyes and can be diluted with culture medium. The PlasMem Bright dyes offer two different color options (green and red) and are provided as ready-to-use DMSO solutions.

Experimental Example: High retentivity on plasma membrane



Fluorescence Properties





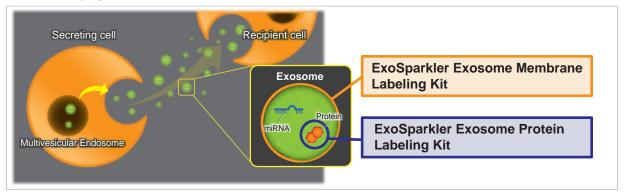
Description	Unit*	Code
PlasMem Bright Green	100 µl	P504-10
PlasMem Bright Red	100 µl	P505-10

Exosome Labeling Kits

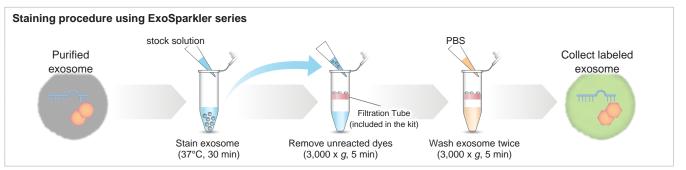


EX10

The ExoSparkler series can be used to stain purified exosomal membrane or protein and allows imaging of labeled exosomes taken up by cells.



Labelling Procedure



ExoSparkler series contains filtration tubes available for the removal of dyes unreacted after fluorescence labeling, as well as an optimized protocol for labeling exosomes. Our ExoSparkler series makes it possible to prepare fluorescence labeling of exosomes using the simple procedure.

Description	Unit	Code
ExoSparkler Exosome Membrane Labeling Kit-Green	5 samples	EX01-10
ExoSparkler Exosome Membrane Labeling Kit-Red	5 samples	EX02-10
ExoSparkler Exosome Membrane Labeling Kit-Deep Red	5 samples	EX03-10
Exosparkler Exosome Protein Labeling Dye-Green	5 samples	EX04-10
Exosparkler Exosome Protein Labeling Dye-Red	5 samples	EX05-10
Exosparkler Exosome Protein Labeling Dye-Deep Red	5 samples	EX06-10
Exolsolator Exosome Isolation Kit	3 test	EX10-10
Exo <i>lsolator</i> Isolation Filter	10 pieces	EX11-10



For inquiries, please contact Dojindo Europe GmbH.

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