

Lipid Peroxidation Probe -BDP 581/591 C11-

Technical Manual

General Information

Lipid peroxides are derived from the oxidation of unsaturated fatty acids by reactive oxygen species. The generated lipid peroxides in turn oxidize lipids, resulting in a chain reaction that increases the amount of lipid peroxides. In addition, when high levels of Fe^{2+} are present, lipid peroxides are generated by the reactions of Fe^{2+} and reactive oxygen species. Lipid peroxides are decomposed into highly reactive aldehydes, such as acrolein, malondialdehyde (MDA), and 4-hydroxy-2-nonenal (4-NHE), which are known to cause cytotoxicity and cell death. Thus, lipid peroxidation causes various diseases, such as atherosclerosis¹⁾.

Lipid Peroxidation Probe -BDP 581/591 C11- is a fluorescent dye used to detect the formation of lipid peroxides. This fluorescent dye does not respond to lipid peroxides, but reacts with lipid radicals formed during lipid peroxidation by reactive oxygen species. Thus, BDP 581/591 C11 can be used to observe the formation of lipid peroxides with the fluorescence color changing from red to green depending on the state of lipid peroxidation. This fluorescent dye allows the detection of the formation of lipid peroxides with high sensitivity because of the high signal-to-noise of the fluorescence signals.

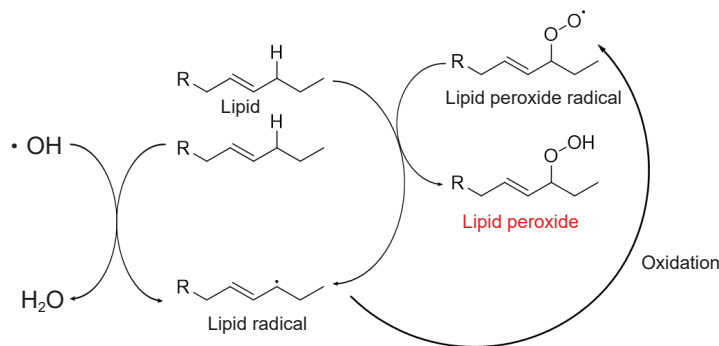


Figure 1. Mechanism of lipid peroxidation

Contents

Lipid Peroxidation Probe -BDP 581/591 C11- 200 tests×1

Storage Conditions

Store in a cool and dark place.

Required Equipment and Materials

- Fluorescence microscope, fluorescence plate reader, or flow cytometer
- Incubator (37°C)
- Micropipettes (100–1000 μl , 20–200 μl , 1–10 μl)
- Microtubes
- Hanks' balanced salt solution (HBSS)
- Dimethyl sulfoxide (DMSO)

Precaution

Centrifuge the tube briefly before opening the cap because the content may be on the tube wall or the ceiling of the cap. Please refer to Table 1 to check suitable fluorescence wavelengths for each application.

Table 1. Recommended filter settings

Applications	Fluorescence plate reader	Fluorescence microscope	Flow cytometer
Fluorescece color: Green	Ex/Em: 480–500/520–540 nm	<ul style="list-style-type: none">• Confocal microscopeEx/Em: 488/510–550 nm• Fluorescence microscopeGFP or FITC filter	FITC filter
Fluorescece color: Red	Ex/Em: 550–570/590–610 nm	<ul style="list-style-type: none">• Confocal microscopeEx/Em: 561/600–630 nm• Fluorescence microscopeTexasRed or Cy3 filter	Cy3 filter

Preparation of Solutions

Preparation of BDP 581/591 C11 DMSO stock solution

Add 20 μl of DMSO to the tube containing Lipid Peroxidation Probe -BDP 581/591 C11- by pipetting. Ensure that the probe is dissolved in the solvent.

Note: Store the reconstituted BDP 581/591 C11 DMSO stock solution at -20°C until use. The solution is stable at -20°C for 1 month.

Preparation of BDP 581/591 C11 working solution

Dilute the BDP 581/591 C11 DMSO stock solution (1:1000) with the culture medium to prepare BDP 581/591 C11 working solution.

Note: The working solution cannot be stored and must be prepared freshly on the day of use. Refer to Table 2 for the preparation of the working solution by vessel type.

Table 2. Required amount of the working solution by vessel type

Vessel	35-mm dish	ibidi 8-well plate	96-well clear black plate (clear bottom)	Flow cytometer
Appropriate amount	2 ml/dish	200 μ l/well	100 μ l/well	0.5 ml/tube

General protocol

- Seed cells on dishes or plates and place them in an incubator at 37°C equilibrated under 95% air and 5% CO₂.
Note: Refer to Table 3 to check the recommended vessels for each detector.

Table 3. Recommended vessels by the detector

Detector	Fluorescence plate reader	Fluorescence microscope	Flow cytometer
Vessel	96-well black plate (clear bottom)	96-well black plate (clear bottom) ibidi 8-well plate 35-mm dish	6-well plate

- Discard the culture medium and wash the cells twice with the culture medium.
- Add an appropriate volume of BDP 581/591 C11 working solution to the cells.
- Incubate the cells as in step 1 for 30 minutes.
- Discard the working solution and wash the cells twice with HBSS.
- Add the culture medium or HBSS containing stimulation agents and incubate at 37°C for an appropriate time.
- Discard the supernatant, wash the cells twice with HBSS, add HBSS, and observe fluorescence signals.

Note: Perform steps 3 and 6 in reverse order if dye staining after drug stimulation is needed.

Usage examples

Detection of lipid peroxidation in HepG2 cells treated with t-BHP (tert-butyl hydroperoxide)

- HepG2 cells (2×10^5 cells/ml) in MEM (supplemented with 10% fetal bovine serum and 1% penicillin-streptomycin) were seeded into vessels and incubated overnight at 37°C under 95% air and 5% CO₂.
Note: (ibidi 8-well plate: 200 μ l/well, 96-well black plate (clear bottom): 100 μ l/well, 6-well plate: 2 ml/well)
- The culture medium was removed, and the cells were washed twice with the culture medium.
- After removing the supernatant, BDP 581/591 C11 working solution was added and the cells were cultured for 30 minutes as in step 1.
Note: (ibidi 8-well plate: 200 μ l/well, 96-well black plate (clear bottom): 100 μ l/well, 6-well plate: 2 ml/well)
- After removing the supernatant, the cells were washed twice with HBSS.
- The supernatant was removed, 200 μ mol/l t-BHP in HBSS was added to each well, and the cells were cultured for 2 hours as in step 1.
Note: (ibidi 8-well plate: 200 μ l/well, 96-well black plate (clear bottom): 100 μ l/well, 6-well plate: 2 ml/well)
- The cells were washed twice with HBSS, and the vessels were refilled with HBSS.
- Fluorescence signals were measured using a plate reader and a flow cytometer, and images were obtained using a fluorescence microscope.

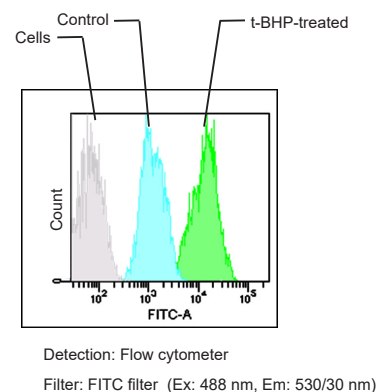
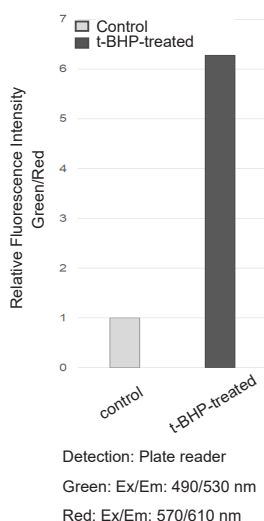
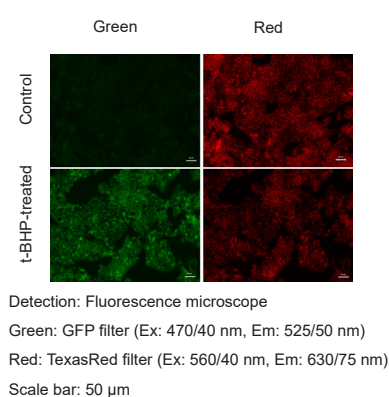


Figure 2. Fluorescence signals from HepG2 cells detected with a fluorescence microscope, flow cytometer, and plate reader

Detection of lipid peroxidation in HepG2 cells treated with cumene hydroperoxide

1. HepG2 cells (2×10^5 cells/ml) in MEM (supplemented with 10% fetal bovine serum and 1% penicillin-streptomycin) were seeded on ibidi 8-well plates and cultured overnight at 37°C under 95% air and 5% CO₂.
2. The culture medium was removed, and the cells were washed twice with the culture medium.
3. After removing the supernatant, BDP 581/591 C11 working solution was added, and the cells were cultured for 30 minutes as in step 1.
4. After removing the supernatant, the cells were washed twice with HBSS.
5. The supernatant was removed, 200 µmol/l cumene hydroperoxide in HBSS was added to each well, and the cells were cultured for 2 hours as in step 1.
6. The cells were washed twice with HBSS, and the vessels were refilled with HBSS.
7. The cells were observed under a fluorescence microscope.

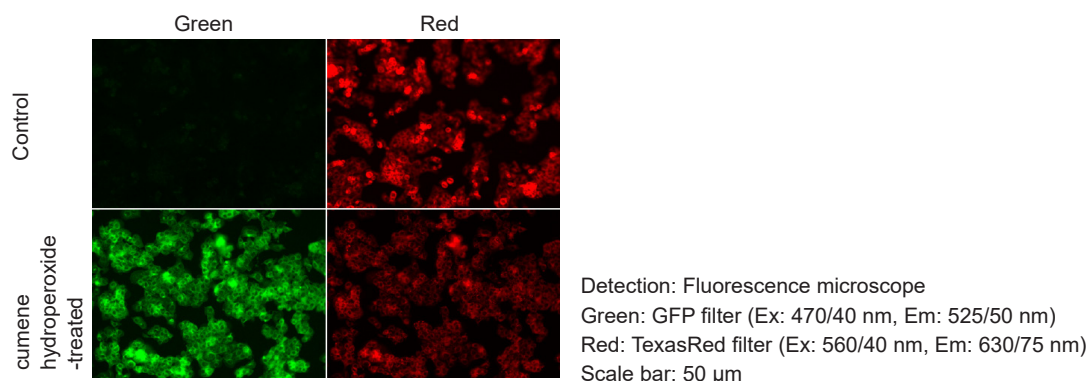


Figure 3. Fluorescent images of HepG2 cells treated with cumene hydroperoxide.

Detection of lipid peroxidation in HepG2 cells treated with Erastin

1. HepG2 cells (2×10^5 cells/ml) in MEM (supplemented with 10% fetal bovine serum and 1% penicillin-streptomycin) were seeded on ibidi 8-well plates and cultured overnight at 37°C under 95% air and 5% CO₂.
2. The culture medium was removed, and the cells were washed twice with the culture medium.
3. The supernatant was removed, 10 µmol/l Erastin in culture medium was added to each well, and the cells were cultured for 24 hours as in step 1.
4. After removing the supernatant, the cells were washed twice with a culture medium.
5. The cells were treated with BDP 581/591 C11 working solution and cultured for 30 minutes as in step 1.
6. After removing the supernatant, the cells were washed twice with HBSS.
7. The cells were observed under a confocal laser scanning microscope.

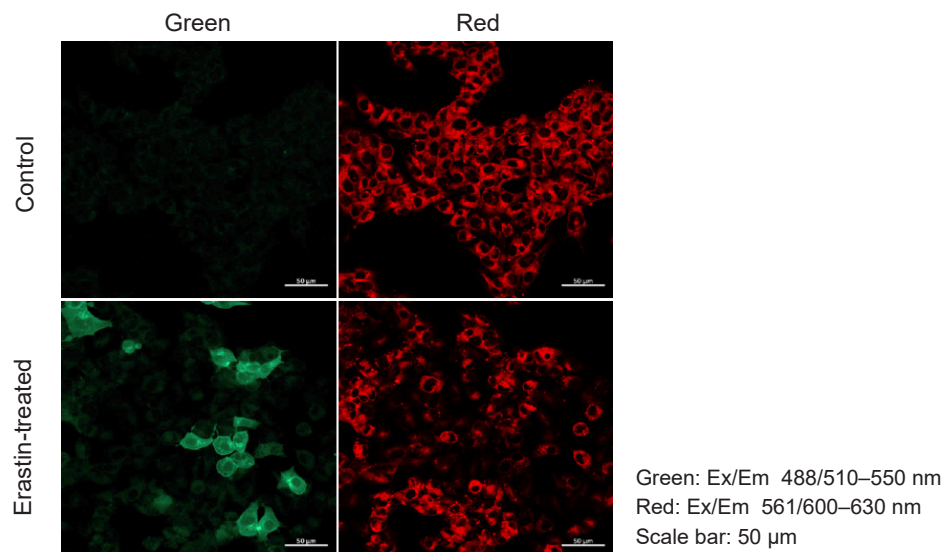


Figure 4. Fluorescent images of HepG2 cells treated with Erastin.

Reference

- 1) S. J. Dixon *et al.*, *Cell*, **2012**, *149*, 1060

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If you need more information, please contact Dojindo technical service.

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